

The Fate of Corals: Will They Overcome Competition with Algae and Cyanobacteria in a Changing Environment?

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Abstract

Coral reefs provide substantial ecological and economic services to coastal communities in the tropics. Hence, there is a great concern about the permanence of these ecosystems and the consequent loss of the economic, ecological and social services coral reefs provide due to their susceptibility to natural and anthropogenic threats. Large-scale processes such as strong El Niño Southern Oscillation events, global warming and ocean acidification represent significant challenges for coral reefs. Benthic cyanobacteria and seaweed have substantially increased in reef areas, facilitated by excessive nutrient input, reduction in herbivore populations and global warming. This review briefly describes the currently known aspects of coral, algae and cyanobacterial interactions, as well as the local and global environmental and ecological aspects that have caused the increase of algae and cyanobacteria in detriment to reef corals. Reef communities will keep changing in the light of large-scale events and anthropogenic influences. As short-term measures, ambitious programs for grazer reintroduction could help curb population growth of algae and cyanobacteria. Medium- to long-term measures should be oriented at limiting nutrient input to water bodies.

Keywords: cyanobacteria, algae, coral reefs, competition, phase changes

1. Introduction

Coral reefs constitute one of the most important, diverse and productive ecosystems in the planet. These ecosystems provide a great number of goods as well as economic and ecological services. Coral reefs protect coastal areas such as seagrasses and mangroves from erosion [1–3].



Coral reefs supply different food sources to coastal communities and also provide habitat for fish, molluscs and crustaceans of commercial interest [4-6]. Coral reefs also harbor a large diversity of microorganisms, algae and invertebrates that have provided researchers, the pharmaceutical, nutraceutical and cosmeceutical industries with many interesting bioactive molecules [7, 8].

Currently, the oceans are suffering remarkable changes evidenced by the increase and intensity of diseases, mass mortalities and blooms of several photosynthetic organisms. All these events have occurred in both disturbed and undisturbed areas around the globe (reviewed in [9]). Among some of the causes that have been linked to these events are global warming, increased UV radiation, overfishing, pollution and coastal eutrophication and oxygen depletion. However, some of all these factors may act in a synergistic manner rather than in isolation, considering that there are large spatial and temporal scales involved in all those processes [9-11].

Degradation of coral reefs has increased considerably world-wide, at least for which there are records of the phenomenon, since there is no consensus of what a pristine reef is [12]; therefore, it is very hard to assess the magnitude of changes and impacts compared to "base line" conditions [13]. Reef degradation has been linked to human activities and impacts; hence, there is a great concern about the permanence of these ecosystems and the consequent loss of the economic, ecological and social services coral reefs provide [14-16]. Moreover, large-scale processes such as the incidence of strong El Niño Southern Oscillation [ENSO] events, global warming and ocean acidification represent an enormous challenge for coral reefs to survive and remain as we know them today [17].

2. Cyanobacteria in reefs

Cyanobacteria are eubacteria with photosynthetic capabilities considered as "primitive microalgae" for a long time. These are ancient organisms dating back to 3.5 billion years as evidenced by their fossil record, particularly in stromatolites [18]. Recent molecular data have revealed that cyanobacteria are a polyphyletic group, and taxa, traditionally grouped by having similar morphologies, have different phylogenetic affiliations [19–22]. Cyanobacteria have a wide distribution in terrestrial and aquatic environments. Some taxa are involved in close symbiotic relationships with diatoms, sponges, corals, lichens and plants such as water ferns and cycads [23-26]. Many cyanobacteria are able to fix atmospheric nitrogen, making a significant contribution of this element into environments where it is usually limiting [27–30].

Marine benthic cyanobacteria thrive in a wide variety of habitats including rocky coastlines, sandy beaches, mangroves, marshes and swamps. The distribution of cyanobacterial mats is influenced by sediment type, tidal exposure and wave energy [31]. In coral reefs, given their abundance at certain times and locations, cyanobacteria may play a similar role to algae in terms of primary production and interactions with herbivores [32]. Cyanobacteria have long been regarded as regular elements of tropical marine communities and for that reason included in floristic lists from a number of tropical regions worldwide [33–35]. However, blooms of marine benthic cyanobacteria have become frequent in coastal areas receiving significant nutrient input from runoff and groundwater, as well as a result of large-scale climatic and hydrological changes [10, 11, 26, 36, 37]. Cyanobacterial blooms, toxin production and derived health risks are currently considered a public health hazard [38–43].

Blooms of marine benthic cyanobacteria grow on the substrate forming dark flimsy mats, usually red, purple, brown or black. There are several reports of marine benthic cyanobacteria blooms in several sites in the Pacific, in the island of Guam [44–46], in some Micronesian atolls and also in the Hawaiian Islands [50]. In the Atlantic, cyanobacterial blooms are a recurrent event strongly subject to seasonality in the state of Florida [51, 52]. In the Caribbean, there are reports from Puerto Rico [53], the Bahamas, Belize and the Virgin Islands [52], and the Colombian Caribbean [35, 54, 55]. Cyanobacterial blooms not only occur in coral reefs but also in seagrass meadows [56–58], mangroves [56, 57], estuaries and coastal lagoons [59].

Blooms in Guam and Australia, due to their extension, persistence and toxicity, have caused mass mortalities of fishes, as well as significant economic losses and health problems in residents or visitors to coastal areas. Marine turtles and manatees are also impacted by these blooms due to toxin exposure [60, 61]. Additionally, benthic cyanobacteria are often the dominant organisms that colonize coral skeletons after bleaching events [62]. Although several species of cyanobacteria grow in microbial mats, these tend to be dominated by a single species, although the reasons by which a species dominates over the other(s) are not yet entirely clear [63]. Additionally, cyanobacteria and turf algae may prevent or inhibit the settlement of coral larvae [50, 64-67]. The increasing occurrence, frequency and intensity of benthic cyanobacterial blooms, is now considered a great threat to coral reefs around the world [51]. Benthic cyanobacteria most commonly linked to bloom formation are included in the orders Chroococcales and Oscillatoriales, with the most common genera being Schizothrix, Hormothamnion, Oscillatoria and Lyngbya [68]. Lyngbya, in particular, is probably the most studied genus for which there is a great number of bloom reports and whose chemistry and chemical ecology are better documented [44, 45, 69-75]. However, recent studies have reassessed the phylogenetic affiliation of this genus finding that it is a rather diverse complex of species now reassigned into the genera Moorea, Okeania or Lyngbya, which may explain the vast chemical complexity of what used to be considered a single genus [19-22]. Although the taxonomy of the group has changed substantially, for the purpose of this review, I will refer to the names that were originally assigned in the papers where they were described.

Blooms of benthic cyanobacteria develop fast, covering vast areas in a period of few weeks. In those events, cyanobacteria grow on the substrate forming mats that can smother benthic organisms. As the bloom progresses, it can turn adjacent sediment and waters anoxic [49]. Cyanobacterial detachment from the sediment, either by flotation or wave action, may promote their dispersion [49, 68].

3. Algae in reefs

Algae are very important members of reef communities where they provide food and refuge to many organisms. They are also the basis of the reef food web [76]. Due to their different phylogenetic affiliations, varying forms and functions, algae are classified into three large functional groups based on growth form and size, ecological characteristics, toughness, susceptibility to herbivores and photosynthetic abilities. Functional groups are used to characterize algal communities in coral reefs and to understand their distribution and responses to environmental factors [76].

Filamentous and small algae with fast growth rates are included in the turf algae category. Multispecies assemblages of turf algae can be very abundant in reefs despite their low biomass. They are an attractive food source for several herbivores, preventing their overgrowth. Cyanobacteria are also included in this category [76]. The next category, fleshy macroalgae, is commonly known as seaweeds and includes the most well-known forms. Seaweed can be very abundant in reefs and have several adaptations to avoid herbivory [77-79]. A third category, coralline or crustose algae, secretes calcium carbonate and is important in cementing the reef framework but also in attracting coral recruits [80, 81].

4. Factors favoring the growth of algae and cyanobacteria in reefs

Competition for light and space between benthic algae and corals affects coral resilience and is essential when assessing the degree of reef health [65, 76]. In healthy coral reefs, reduced nutrient availability and high grazing pressure are the most important factors controlling the growth of algal turf and fleshy algae [82]. Phase shifts or phase changes are evident by a decrease in the cover and recruitment of corals compared to the colonization and substrate cover by turf algae, seaweed and cyanobacteria [14, 65, 83]. Phase shifts are common in many degraded reefs due to disparity in coral-algal interactions [15, 76, 86]. Under stressful conditions, favored by bleaching events, partial mortality or reduced grazing pressure, algae and cyanobacteria become competitively superior to corals, eventually overgrowing and killing them [15, 87].

Alleged causes linked to cyanobacterial blooms in fresh and brackish waters have been thoroughly studied (reviewed in [88]). However, blooms of marine benthic cyanobacteria are not understood that well. Normally, they occur in shallow waters, and environmental factors such as high temperatures, reduced wave action and the availability of phosphorus, nitrogen and iron have been linked to their formation [48, 49, 66, 89, 90]. Individual strains of cyanobacteria vary greatly in their bloom dynamics, which in some cases may be controlled by physical disturbances rather that nutrient availability or competition with macroalgae [91]. Several studies, however, have established correlations between bloom formation and mild wave action and increases in water temperature, phosphate levels and/or iron bioavailability. Bloom persistence, however, has been correlated to the low palatability of cyanobacteria to most generalist herbivores [45, 50, 70, 88, 92, 93].

Algal and cyanobacterial blooms may be difficult to explain because as multispecies consortia (at least in the case of cyanobacterial blooms) the dynamics in bloom formation may be a result

of species-specific adaptations or preferences (reviewed in [94]). Also, the resulting increased growth may be a result of nutrient loading and/or reduction or absence of grazing [94]. Hence, food web dynamics may be important to understand this phenomenon. However, it is a nobrainer that the removal of large and small herbivores [12] has certainly favored the growth of cyanobacteria and algae, which have taken advantage of the increased nutrient availability and decreased grazing pressure.

5. Water temperature and cyanobacterial blooms

Sustained water temperatures over 28°C, alterations of flow and time of residence, decrease in water viscosity, large-scale mixing processes and small-scale turbulence are physical factors that correlate with cyanobacterial bloom formation [10, 88, 89, 95].

Thorough monitoring of *Lyngbya majuscula* blooms in the coasts of Puerto Rico from September 1998 until February 2000 showed that during September 1998 and September 2000 to December 2000, *Lyngbya* achieved the greatest cover coinciding with the highest temperatures recorded. While mean cover values ranged between 7 and 82% year-round, *Lyngbya* cover reached up a 100% during the warmest periods [53]. In Rosario Islands, Colombia, a similar pattern was observed in September 2010 and 2011, where the cover of cyanobacterial mats reached a 100%, concurring with sustained water temperatures above 30°C [93].

In Australia, benthic cyanobacterial blooms are recognized as an environmental health hazard [47, 49, 96]. Monitoring of L. majuscula mats in eastern Australia during 2005 showed that during the cool winter months blooms did not develop, but between the months of October and November, when water temperature increased, there was a fast development of *Lyngbya* mats. Blooms began gradually at the end of October, when total bottom cover of L. majuscula did not surpass 10% in an area of 49 hectares. During November, the total bottom cover of L. majuscula reached values close to 40% in an approximate area of 329 hectares. At the same time, water temperature increased from 23.5°C in October to 30.8°C in December. By the end of December, the bloom covered almost a 100% of the total available substrate in an area of 529 hectares [89]. I calculated an approximate wet biomass of 5000 Tons when the bloom reached its peak. As temperature decreased, so did Lyngbya cover. Similar patterns were registered in the Colombian Caribbean. Mats of L. majuscula and L. sordida covered extensions close to a 100% of the available reef substrate in Rosario Islands in September 2010 when the water temperature surpassed 30°C [93]. Also, mats of Symploca hydnoides and Phormidium submembranaceum were very conspicuous in Old Providence Island during October 2009 and July 2010. Again, those blooms took place during the warmest months recorded in the San Andres Archipelago [above 28°C] [55, 93].

6. Nutrient input favors the growth of algae and cyanobacteria

Water quality degradation as a result of increased nutrient input promotes the development and persistence of algal blooms and is one of the reasons that explains their expansion worldwide [94]. Although assessing the influence of nutrients is far more complicated than correlating these with increased water temperatures, experts agree that nutrients play a significant role in favoring blooms. Regular or pulsated nutrient delivery promotes bloom development; the overall composition—not just concentration—of nutrient input impacts blooms and high-biomass blooms may be partly sustained by exogenous nutrient sources [94].

Several studies have shown that cyanobacterial blooms are stimulated by the increase in nutrient concentration in water bodies. Nutrient increase may be a result of sewage disposal, animal husbandry or agricultural activities [26, 41, 49, 56, 57, 88, 89, 95, 97]. Rains also stimulate bloom formation due to increased runoff and river flow providing extra amounts of nutrients from terrestrial environments to coastal areas [49, 55, 98]. Groundwater and atmospheric deposition may also influence bloom formation and dynamics [94]. Nutrients that do have a stimulating growth effect on cyanobacteria are iron, phosphorus and nitrogen [26, 36, 49, 57, 89, 90, 99]. In the Colombian Caribbean, I have detected an apparent synergistic effect of high temperatures, the onset of rainy seasons and nutrient increase with incidence of marine cyanobacteria blooms at various sites [55, 93].

Anthropogenic activities such as the use of fertilizers and the dumping of urban and industrial waste waters provide significant amounts of nutrients into water bodies [26]. The input of key nutrients favor the growth of phototrophic organisms such as plants, algae and cyanobacteria, largely controlling aquatic primary production in marine environments where nitrogen is usually limiting. These blooms, with oxygen consumption during the hours of darkness, generate anoxia and consequent mortality of fish and other aquatic organisms [56].

7. The effect of cyanobacteria and algae on corals

There is significant evidence that reef degradation has increased on a global scale. Just in the Caribbean region, at least 20% of live coral coverage has been lost per decade [6]. This loss has been attributed to the increase in human population, dumping of waste water, soil erosion and subsequent sediment input by rivers, eutrophication, imbalance of food chains, proliferation of macroalgae [65, 100, 101], diseases and climate change [101, 102], as well as the removal or decimation of top predators and large herbivores [12].

Competition is a process that determines the structure, composition and diversity of benthic communities in coral reefs [5, 103–107]. In coral reefs, competition between sessile organisms such as benthic algae and corals has become very significant for the resilience of corals due to the dominance and vast cover of algae and cyanobacteria in affected coral reefs [65]. Reef deterioration has caused a significant replacement of live coral cover by cyanobacteria and macroalgae [59, 83, 108].

The strong competition for space between coral, algae and marine cyanobacteria can determine the structure, composition and abundance of these three groups in coral reefs [59, 109]. Overgrowth of algae or cyanobacteria on corals can cause deleterious effects on their health [50, 54, 55, 66, 110]. The vast occurrences of benthic algae and cyanobacteria in coral reefs are certainly an indication of the prevalent ecological conditions and may serve as indicators of coral reef health and local ecological imbalances [51, 111].

Coral reefs have a great diversity of herbivores that can exert a strong pressure on communities of primary producers such as macroalgae and cyanobacteria [2, 15]. In tropical areas, the most important herbivores in terms of consumption and impact on macroalgae are fish and sea urchins [70, 72, 77]. These herbivores can consume between 60 and 100% of the algal biomass daily [112–114]. Herbivory and nutrient input are two more determinant factors that define the relationship between algae and corals in reefs [85, 115] and the success of algae in other coastal ecosystems [116].

However, the generalized decrease in the populations of reef herbivores is an indirect cause of the increase in algal coverage and consequent phase shifts [2, 15, 50, 83]. Overfishing in reef areas has decimated the populations of herbivorous fish causing imbalances in the populations of macroalgae and their consequent overgrowth on corals and other substrates [2, 83, 84]. Occasionally, other herbivores such as sea urchins can sometimes increase in response to the decrease in herbivorous fishes and control algal populations. In the Caribbean basin, however, mass mortalities of sea urchins, particularly *Diadema antillarum*, triggered an excessive increase of algae in Jamaica and other locations [83].

Benthic cyanobacteria are efficient colonizers in a wide array of substrates such as coral skeletons, live coral, sand and even macroalgae. Cyanobacteria increase the magnitude of the phase shifts [51]. Benthic cyanobacteria, as well as macroalgae, are favored by bottom-up (increase in nutrients in the water bodies) and top-down effects (such as the decline of herbivores in reefs) [85]. Various studies have shown that the most important factor in controlling algae populations in coral reef areas is herbivory and the lack of it increases the consequences of phase changes more so than eutrophication, especially in the Caribbean [117].

Seaweed or macroalgae, compared to terrestrial plants, are mostly foliage, lacking dense structural material such as lignin, cellulose and hemicellulose, being more susceptible to herbivore consumption [118]. Benthic cyanobacterial mats strongly resemble macroalgae because they may exhibit large biomass and could become a potential food source for reef herbivores [44]. During blooms, cyanobacteria can surpass macroalgae in terms of available biomass [44, 72, 93].

Herbivory in reef areas generates a strong pressure on macroalgae and cyanobacteria. In "healthy" reefs, grazing rates may reach 100% of the produced biomass [77, 85, 112, 119]. Macroalgae and cyanobacteria, however, minimize grazing by means of spatial or temporal escapes, tolerating herbivory by compensating tissue loss with fast growth rates or investing in structural or chemical defenses [77–79, 113]. The production of feeding deterrent compounds, in particular, is well documented in these groups and plays an important role against grazing, enabling the growth and persistence of algae and cyanobacteria in coral reef areas [78, 119]. Besides deterring herbivores, chemically defended cyanobacteria could be favored by selective fish and urching grazing over palatable macroalgae, removing potential competitors and favoring their growth and expansion in reef areas [26].

Massive colonization of hard substrates by algae and benthic cyanobacteria can determine competitive networks, structure, composition and abundance of these three groups locally [51, 109]. In events where the abundance of cyanobacteria or macroalgae greatly increases, there is also an increase in overgrowth interactions with corals. The overgrowth of cyanobacteria and

algae over corals may cause deleterious effects on their health [50, 66, 110]. In the Colombian Caribbean, the growth of benthic cyanobacterial mats over different species of scleractinian corals and soft corals eventually results in the death of the coral tissue that underlies those mats [54, 93] (**Figure 1**). In the Florida Keys, [50] determined that the brown seaweeds *Dictyota* and *Lobophora*, as well as several species of *Lyngbya*, not only inhibited the larval settlement of the hard coral *Porites astreoides* and the soft coral *Briareum asbestinum*, but also showed increased recruit mortality when the larvae came into direct contact with these organisms. Hence, seaweed and cyanobacteria may be considered competitively superior to corals not just by taking up space but also by inhibiting their recruitment.

Competition through allelopathic mechanisms by benthic marine cyanobacteria causes harmful effects on corals and soft corals. This has been observed in the field and tested in laboratory experiments in various sites such as the Bahamas, Belize, Florida and Panama [52], Florida [120], Hawaii [121] and the Colombian Caribbean [54, 93]. Experiments have been performed over coral embryos and larvae [50, 66] or adult corals [110].

[66] compared the recruitment and survival of embryos of the hard corals *Pocillopora damicornis* and *Acropora surculosa*, in the presence of the cyanobacterium *L. majuscula*. Coral embryos got tangled in the dense Lyngbya filaments and died. Cyanobacterial mats may trap coral embryos but also promote sediment deposition and accumulation surrounding benthic microbial mats [122]. Under these mats, anoxic conditions may develop, which may favor nitrogen fixation but are potentially deadly to the very sensitive coral embryos [66]. Since recruitment is a key process in the maintenance and recovery of coral reef ecosystems [50, 123], any alteration in this process has negative implications and will affect the persistence and resilience of corals.

The fact that allelopathy is so hard to prove experimentally does not mean that it does not occur in reef environments. Cyanobacteria, in particular, while being in direct contact with corals, could release allelopathic compounds as a result of abrasion by water motion with consequent cell rupture. Many studies have shown that toxin release from cyanobacteria is a result of cell lysis due to abrasion, stress and cell death [124–126]. Additionally, environmental factors such as temperature may elicit active toxin liberation in these microorganisms [124, 126].

Competitive interactions between the hard coral *Porites lutea*, a brown alga *Dictyota dichotoma* and the cyanobacterium *Lyngbya bouillonii* were evaluated in Sesoko Island, Japan. While coral growth is compromised by direct contact and abrasion by the alga, mats of the cyanobacterium *L. bouillonii* are able to kill live coral tissue upon direct contact [110].

Interactions between cyanobacteria, a hard coral and a soft coral in Rosario Islands, Colombian Caribbean, were evaluated *in situ* in order to identify deleterious effects potentially related to allelopathic mechanisms. Cyanobacterial extracts were incorporated into Phytagel™ gels and these were placed in direct contact with the hard coral *Porites porites* and the soft coral *B. asbestinum* [127]. HPLC chromatographic profiles of zooxanthellae in coral tissues were evaluated after 24, 48 and 72 hours of exposure. Extracts from *Lyngbya* spp. showed a clear effect on the zooxanthellae chromatographic profiles evidenced by an increase in pheophytin, a degradation product from chlorophyll. The effect was greater with longest exposure time. These results suggest that cyanobacteria may compete against corals due to their fast growth rates, defenses against herbivory and allelopathic mechanisms. Further evidence was

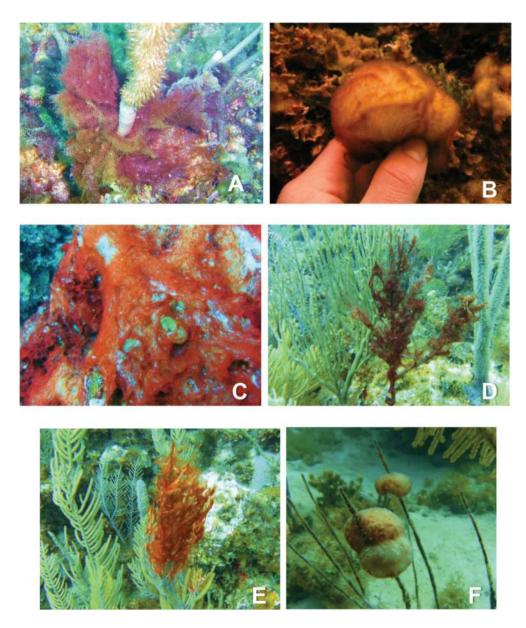


Figure 1. Benthic cyanobacteria from Colombian Caribbean reefs. (A) Mats of *Moorea producens* growing at the base of a *Eunicea* soft coral. The soft coral retracts its polyps due to abrasion and possibly to avoid contact with the mat. Rosario Islands, 2016. (B) Filamentous mats of *Caldora* sp. This cyanobacterium grows profusely over various kinds of seaweeds. Old Providence Island, 2008. (C) Unidentified red filamentous mats overgrowing hard corals and green algae. Old Providence Island, 2008. (D and E) Multispecies cyanobacterial mats overgrowing soft corals, causing polyp necrosis and issue death. The remaining gorgonin skeletons continue to be colonized by other cyanobacteria and filamentous algae. Old Providence Island, 2016. (F) Puffs of filamentous cyanobacteria may overgrow live corals causing bleaching and/or necrosis. In this case, they are growing over gorgonin skeletons. Old Providence Island, 2016. Photos A, D–F: Monica Puyana; B–C: Julian Prato.

obtained while testing the potential allelopathic effects of two organic extracts from marine cyanobacteria over live corals. Organic extracts from two different microbial consortia, a consortium of *S. hydnoides* and *P. submembranaceum* and a consortium of *Lyngbya cf. semiplena*, *L. majuscula*, *Oscillatoria nigroviridis* and *O. margaritifera*, were tested at 0.002 ppm and 0.011 ppm, both below their natural concentration. In order to do so, fragments of the hard coral *Madracis mirabilis* were placed in individual containers, and once acclimated, with their polyps fully expanded, cyanobacterial extracts were resuspended in ethanol and seawater and coral behavior was registered. Both extracts generated the retraction of coral polyps with variable speed and intensity. Ethanol controls, on the other hand, presented a rapid recovery and a minimum shrinkage of polyps. Fragments exposed to cyanobacterial extracts showed 80 to 99% of polyp retraction, whereas solvent controls did not exceed 25% of polyp retraction. No extract proved to be lethal, and after 20 hours, corals extended their polyps displaying full recovery [93].

In another assay, the toxicity of four cyanobacterial extracts to embryos of *Montastrea annularis*, obtained during the mass spawning event in Rosario Islands in September, 2011, was evaluated. In this assay, we assessed embryo mortality 6, 18 and 24 hours after acute exposure to cyanobacterial extracts in concentrations of 1000, 500, 100 and 10 ppm. All tested extracts were toxic at concentrations of 500 and 1000 ppm, causing 100% embryo mortality within 6 hours. Controls with seawater and ethanol did not affect coral embryos. The extract that showed greater toxicity was obtained from a mixed *Lyngbya* assemblage, causing a 90% embryo mortality at the lowest concentration tested (10 ppm) after only 6 hours of exposure.

In summary, competition between corals, cyanobacteria and/or algae may take place either against coral embryos or larvae or during their adult stages. Some of the mechanisms include allelopathic inhibition on adult corals and embryos, negative effects of abrasion by direct physical contact, drastic decrease in oxygen levels near and under cyanobacterial mats, reduction of available space in reef substrates affecting larval recruitment and mortality of embryos entangled in cyanobacterial mats. All these effects strongly suggest that blooms of algae and benthic cyanobacteria pose a risk for the recruitment and development of reef builders and other reef organisms such as soft corals [50, 51, 66, 93, 110, 120, 121].

8. Climate change, ocean acidification and future of reefs

Resilience or the ability to withstand and recover from the negative phenomena affecting coral reefs is an essential role for their persistence. Recovery after each disturbance involves the re-establishment of coral coverage, the growth of surviving coral fragments, reproductive success and the subsequent settlement and survival of coral embryos and larvae [16, 123]. However, the recovery process in coral reefs is nowadays more difficult because the substrate in these ecosystems is frequently dominated by algae and cyanobacteria. These photosynthetic organisms efficiently colonize available substrates following disturbances and become dominant in degraded reefs [15, 65, 83, 123], reducing the resilience of coral reefs overall

[2, 15]. Algal dominance can become a serious bottleneck for the re-establishment of corals, depending on the characteristics of the dominant algal assemblages in each place [65, 123].

Large-scale hydrological disturbances and environmental changes due to global warming and ocean acidification are recognized as growing threats to coral reefs worldwide [17, 123, 128–130]. Global warming is also believed to facilitate the development, frequency and expansion of cyanobacterial blooms [10, 11, 26, 37]. These large-scale disturbances may have a more direct effect on corals, either by reducing growth rates and calcification regimes under acidic seawater conditions or by the onset of bleaching events under warmer conditions [17, 123]. Unless there is a significant reduction in CO₂ atmospheric levels, reefs with reduced populations of grazers and increased nutrient input are predicted from shift from predominantly coral-dominated to predominantly algal-dominated states [17, 130]. This pattern has been recognized for some time now, particularly in the Caribbean and the Eastern Pacific [131–135]. Management options such as restoring herbivore populations and limiting nutrient input to reef areas may only be effective under controlled CO, input to seawater [17, 123, 130]. Sea urchins are important grazers of noxious algae [136]; therefore, restoration of grazers such as urchins is believed to alleviate the problem of excessive algal biomass in coral reefs. In Hawaii, juveniles of the local urchin Tripneustes gratilla are raised in hatcheries and reintroduced in reef areas in order to remove excessive biomass of invasive algal species and restore the natural ecosystem function [137]. However, control of cyanobacterial populations by grazers seems very unlikely [26].

Approximately two-thirds of coral reefs in the Caribbean are threatened by human activities such as coastal development, wastewater and sediment input, pollution and overfishing [138]. The economic impact of coral bleaching, coral diseases and cyanobacterial blooms has not been quantified, but it is clear that these have caused significant changes in Caribbean reef communities [138]. Therefore, it is important to assess the impact of cyanobacterial blooms at local scales in order to understand their causes and consequences in order to address significant monitoring and management measures. The relationships between nutrient input and bloom formation are obscured by changes in food webs, habitat alterations and climate change. Nutrient enrichment has several effects in food webs, predator-prey interactions and overall nutrient dynamics. However, experts agree that management of nutrient inputs to watershed may be the most important measure to prevent excessive growth of algae and cyanobacteria [26, 94]. The Australian government, for instance, has invested significant funds to reduce nitrogen inputs into Moreton Bay to prevent blooms of *L. majuscula* [90].

9. Conclusions

In order to answer the question whether corals will overcome competition with algae and cyanobacteria in environments, the evidence shows that reef communities have changed over time and will keep changing in the light of large-scale events and anthropogenic influences. Algae and cyanobacteria are thriving as the total bottom cover of reef corals is reduced. So even if corals survive, their communities will not be the same. We do need to gain a better

understanding of causes, at local and regional scales, that support bloom formation. Also, it is crucial to convince managers that only long-term studies and periodic surveys will help to understand and manage algae and cyanobacteria in reefs and other marine ecosystems. As short-term measures, ambitious programs for grazer reintroduction could help curb population growth of algae and cyanobacteria. Medium- to long-term measures should be oriented at limiting nutrient input to water bodies.

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References

- [1] Moberg F, Folke C. Analysis: Ecological goods and services of coral reef ecosystems. Ecological Economics. 1999;29:215-233
- [2] Hughes TP, Baird AH, Bellwood DR, Card M, Connolly SR, Folke C, Grosberg R, Hoegh-Guldberg O, Jackson JBC, Kleypas J, Lough JM, Marshall P, Nyström M, Palumbi SR, Pandolfi JM, Rosen B, Roughgarden J. Climate change, human impacts, and the resilience of coral reefs. Science. 2003;301:929-933
- [3] Ritson-Williams R, Arnold SN, Fogarty ND, Steneck RS, Vermeij MJA, Paul VJ. New perspectives on ecological mechanisms affecting coral recruitment on reefs. In: Lang MA, MacIntyre IG, Rützler K, editors. Proceedings of the Smithsonian Marine Science Symposium. Vol. 2009. Washington, DC: Smithsonian Institution Scholarly Press; 2009. p. 437-457
- [4] Jennings S, Polunin NVC. Impacts of fishing on tropical reef ecosystems. Ambio. 1996; 25:44-49
- [5] Birkeland C. Life and Death of Coral Reefs. New York: Chapman and Hall; 1997. 536 p
- [6] Mumby PJ, Steneck R. Coral reef management and conservation in the light of rapidly evolving ecological paradigms. Trends in Ecology and Evolution. 2008;23:555-563
- [7] Molinski TF, Dalisay DS, Lievens SL, Saludes JP. Drug development from marine natural products. Nature Reviews-Drug Discovery. 2009;8:69-85
- [8] Leal MC, Puga J, Serôdio J, Gomes NCM, Calado R. Trends in the discovery of new marine natural products from invertebrates over the last two decades-where and what are we bioprospecting? PLoS One. 2012;7:e30580. DOI: 10.1371/journal.pone.0030580
- [9] Hayes ML, Bonaventura J, Mitchell TP, Prospero JM, Shinn EA, Van Dolah F, Barber RT. How are climate and marine biological outbreaks functionally linked? Hydrobiologia. 2001;460:213-220. DOI: 10.1023/A:1013121503937
- [10] Paerl HW, Huisman J. Blooms like it hot. Science. 2008;320:57-58
- [11] Paerl HW, Huisman J. Climate change: A catalyst for global expansion of harmful algal blooms. Environmental Microbiology Reports. 2009;1:27-37
- [12] Jackson JBC. What was natural in the coastal oceans? Proceedings of the National Academy of Sciences. 2001;98:5411-5418
- [13] Bruno JF, Precht WF, Vroom PS, Aronson RB. Coral reef baselines: How much macroalgae is natural? Marine Pollution Bulletin. 2014;80:24-29. DOI: 10.1016/j.marpolbul.2014.01.010
- [14] Pandolfi JM, Bradbury RH, Sala E, Hughes TP, Bjorndal KA, Cooke RG, McArdle D, McClenachan L, Newman MJH, Paredes G, Warner RR, Jackson JBC. Global trajectories of the long-term decline of coral reef ecosystems. Science. 2003;301:955-958

- [15] Hughes TP, Rodrigues MJ, Bellwood DR, Ceccarelli D, Hoegh-Guldberg O, McCook L, Moltschaniwskyj N, Pratchett MS, Steneck RS, Willis B. Phase shifts, herbivory, and the resilience of coral reefs to climate change. Current Biology. 2007;17:360-365
- [16] Birrell CL, McCook LJ, Willis BL, Diaz-Pulido GA. Effects of Benthic algae on the replenishment of corals and the implications for the resilience of coral reefs. Oceanography and Marine Biology: An Annual Review. 2008;46:25-63
- [17] Anthony KRN, Maynard JA, Diaz-Pulido G, Mumby PJ, Marshall PA, Cao L, Hoegh-Guldberg O. Ocean acidification and warming will lower coral reef resilience. Global Change Biology. 2011;17:1798-1808. DOI: 10.1111/j.1365-2486.2010.02364.x
- [18] Whitton BA, Potts M. Introduction to the Cyanobacteria. In: Whitton BA, Potts M, editors. The Ecology of Cyanobacteria. Dordrecht: Kluwer Academic Publishers; 2000. p. 1-11
- [19] Engene N, Coates RC, Gerwick WH. 16S rRNA gene heterogeneity in the filamentous marine cyanobacterial genus Lyngbya. Journal of Phycology. 2010;46:591-601
- [20] Engene N, Choi H, Esquenazi E, Rottacker E, Ellisman MH, Dorrestein PC, Gerwick WH. Underestimated biodiversity as a major explanation for the perceived rich secondary metabolite capacity of the cyanobacterial genus Lyngbya. Environmental Microbiology. 2011;13:1601-1610
- [21] Engene N, Rottacker EC, Kasvtovsky J, Byrum T, Choi H, Ellisman MH, Komárek J, Gerwick WH. Moorea producens gen. nov., sp. nov. and Moorea bouillonii comb. nov., tropical marine cyanobacteria rich in bioactive secondary metabolites. International Journal of Systematic and Evolutionary Microbiology. 2012;62:1171-1178
- [22] Engene N, Paul V, Byrum T, Gerwick W, Thor A, Ellisman M. Five chemically rich species of tropical marine cyanobacteria of the genus Okeania gen. nov. [Oscillatoriales, Cyanoprokaryota]. Journal of Phycology. 2013;69:1095-1106
- [23] Adams DG. Cyanobacteria in symbiotic interactions. In: Whitton BA, Potts M, editors. The Ecology of Cyanobacteria. Vol. 2000. Dordrecht: Kluwer Academic Publishers; 2000. p. 523-561
- [24] Cox PA, Banack SA, Murch SJ. Biomagnification of cyanobacterial neurotoxins and neurodegenerative disease among the Chamorro people of Guam. Proceedings of the Natural Academy of Sciences. 2003;100:13380-13383
- [25] Usher KM. The ecology and phylogeny of cyanobacterial symbionts in sponges. Marine Ecology. 2008;29:178-192
- [26] Paerl HW, Paul VJ. Climate change: Links to global expansion of harmful cyanobacteria. Water Research. 2012;49:1349-1363
- [27] Corredor JE, Wilkinson CR, Vicente VP, Morell JM, Otero E. Nitrate release by Caribbean reef sponges. Limnology and Oceanography. 1988;33:114-120
- [28] Capone DG, Zehr JP, Paerl HW, Bergman B, Carpenter EJ. Trichodesmium, a globally significant marine cyanobacterium. Science. 1997;276:1221-1229

- [29] Diaz MC, Ward BB. Sponge mediated nitrification in tropical benthic communities. Marine Ecology Progress Series. 1997;156:97-107
- [30] Ferris MJ, Palenik B. Niche adaptation in ocean cyanobacteria. Nature. 1998;396:226-228
- [31] Hoffman L. Marine cyanobacteria in tropical regions: Diversity and ecology. European Journal of Phycology. 1999;34:371-379
- [32] Paul VJ, Cruz-Rivera E, Thacker R. Chemical mediation of macroalgal-herbivore interactions. In: McClintock J, Baker B, editors. Marine Chemical Ecology. Boca Raton: CRC Press; 2001. p. 227-265
- [33] Littler DS, Littler MM. 2000. Caribbean Reef Plants. An Identification Guide to the Reef Plants of the Caribbean, Bahamas, Florida and Gulf of Mexico. Offshore Graphics: Washington; 2000. 542 p
- [34] Diaz-Pulido G, Diaz-Ruiz M. Diversity of benthic marine algae of the Colombian Atlantic. Biota Colombiana. 2003;4:203-246
- [35] Reyes V, Gavio B, Velasquez H. Notes on the marine algae of the International Biosphere Reserve Seaflower, III. New records of Cyanophyta for the Caribbean coast of Colombia. Nova Hedwigia. 2013;97:349-360
- [36] Ahern KS, Ahern CR, Udy JW. In situ field experiment shows *Lyngbya majuscula* [cyanobacterium] growth stimulated by added iron, phosphorus and nitrogen. Harmful Algae. 2008;7:389-404
- [37] Paerl HW, Fulton RS. Ecology of harmful cyanobacteria. In: Graneli E, Turner J, editors. Ecology of Harmful Marine Algae. Berlin: Springer-Verlag; 2006. p. 95-107
- [38] Paerl HW. A comparison of cyanobacterial bloom dynamics in freshwater, estuarine and marine environments. Phycologia. 1996;35:25-35
- [39] Sivonen K. Cyanobacterial toxins and toxin production. Phycologia. 1996;35(6 suppl): 12-24
- [40] Sellner KG. Physiology, ecology, and toxic properties of marine cyanobacterial blooms. Limnology and Oceanography. 1997;42(5, part 2):1089-1104
- [41] Metcalf JS, Codd GA. Cyanobacterial Toxins in the Water Environment: A Review of Current Knowledge. UK: Foundation for Water Research. 2004. 36 p
- [42] Miller MA, Kudela RM, Mekebri A, Crane D, Oates SC, Tinker MT, Staedler M, Miller WA, Toy-Choutka S, Dominik C, Hardin D, Langlois G, Murray M, Ward K, Jessup DA. Evidence for a novel marine harmful algal bloom: Cyanotoxin (microcystin) transfer from land to sea otters. PLoS One. 2010;5:e12576. DOI: 10.1371/journal.pone.0012576
- [43] Leao PN, Engene N, Antunes A, Gerwick WH, Vasconcelos V. The chemical ecology of cyanobacteria. Natural Products Reports. 2012;29:372-391
- [44] Pennings S, Pablo SR, Paul VJ. Chemical defenses of the tropical, benthic marine cyanobacterium *Hormothamnion enteromorphoides*: Diverse consumers and synergisms. Limnology and Oceanography. 1997;**42**:911-917

- [45] Nagle D, Paul VJ. Chemical defense of a marine cyanobacterial bloom. Journal of Experimental Marine Biology and Ecology. 1998;225:29-38
- [46] Thacker RW, Ginsburg DW, Paul VJ. Effects of herbivore exclusion and nutrient enrichment on coral reef macroalgae and cyanobacteria. Coral Reefs. 2001;19:318-329
- [47] Dennison WC, O'Neil JM, Duffy EJ, Oliver PE, Shaw GR. Blooms of the cyanobacterium Lyngbya majuscula in coastal waters of Queensland, Australia. Bulletin de l'Institut océanographique [Monaco]. 1999;19:501-506
- [48] O'Neil JM, Shaw GR, Dennison WC. Blooms of the toxic cyanobacteria Lyngbya majuscula in coastal Queensland waters. In: Abstracts 9th International Conference on Algal Blooms. Tasmania, 2000
- [49] Albert S, JM O'N, Udy JW, Ahern KS, CM O'S, Dennison WC. Blooms of the cyanobacterium Lyngbya majuscula in coastal Queensland, Australia: Disparate sites, common factors. Marine Pollution Bulletin. 2005;51:428-437
- [50] Kuffner IB, Walters LJ, Becerro MA, Paul VJ, Ritson-Williams R, Beach KS. Inhibition of coral recruitment by macroalgae and cyanobacteria. Marine Ecology Progress Series. 2006;323:107-117
- [51] Paul VJ, Thacker R, Banks K, Golubic S. Benthic cyanobacterial blooms impact the reef of south Florida [Broward County, USA]. Coral Reefs. 2005;24:693-697
- [52] Ritson-Williams R, Paul VJ, Bonito V. Marine benthic cyanobacteria overgrow coral reef organisms. Coral Reefs. 2005;24:629
- [53] Stielow S, Ballantine D. Benthic cyanobacterial, Microcoleus lyngbyaceus, blooms in shallow, inshore Puerto Rican seagrass habitats, Caribbean Sea. Harmful Algae. 2003;2:127-133. DOI: 10.1016/S1568-9883(03)00007-6
- [54] Puyana M, Prato J. Overgrowth of reef organisms by benthic cyanobacteria in the Colombian Caribbean. Mutisia. 2013;3:58-60
- [55] Puyana M, Acosta A, Bernal-Sotelo K, Velásquez-Rodríguez T, Ramos F. Spatial scale of cyanobacterial blooms in Old Providence Island, Colombian Caribbean. Universitas Scientiarum. 2015;20:83-105. DOI: 10.11144/Javeriana.SC20-1.sscb
- [56] Paerl HW. Controlling eutrophication along the freshwater-marine continuum: Dual nutrient (N and P) reductions are essential. Estuaries and Coasts. 2009;32:593-601
- [57] Pittman SJ, Pittman KM. Short-term consequences of a benthic cyanobacterial bloom (Lyngbya majuscula Gomont) for fish and penaeid prawns in Moreton Bay [Queensland, Australia]. Estuarine, Coastal and Shelf Science. 2005;63:619-632
- [58] Conley D, Paerl H, Howarth R, Boesch D, Seitzinger S, Havens K, Lancelot C, Likens G. Controlling eutrophication: Nitrogen and phosphorus. Science. 2009;323:1014-1015. DOI: 10.1126/science.1167755
- [59] Charpy L, Casareto B, Langlade M-J, Suzuki Y. Cyanobacteria in coral reef ecosystems: A review. Journal of Marine Biology. 2012;2012:1-9. DOI: 10.1155/2012/259571

- [60] Arthur KE, Limpus CJ, Roelfsema CM, Udy JW, Shaw GR. A bloom of *Lyngbya majuscula* in Shoalwater Bay, Queensland, Australia: An important feeding ground for the green turtle (*Chelonia mydas*). Harmful Algae. 2006;**5**:251-265
- [61] Harr KE, Szabo NJ, Cichra M, Phlips EJ. Debromoaplysiatoxin in *Lyngbya*-dominated mats on manatees (*Trichechus manatus latirostris*) in the Florida King's Bay ecosystem. Toxicon. 2008;52:385-388
- [62] Diaz-Pulido G, McCook LJ. The fate of bleached corals: Patterns and dynamics of algal recruitment. Marine Ecology Progress Series. 2002;232:115-128
- [63] Stal LJ. Physiological ecology of cyanobacteria in microbial mats and other communities. The New Phytologist. 1995;131:1-32
- [64] Edmunds PJ, Carpenter RC. Recovery of *Diadema antillarum* reduces macroalgal cover and increases abundance of juvenile corals on a Caribbean reef. Proceedings of the National Academy of Sciences of the United States of America. 2001;98:5067-5071. DOI: 10.1073/pnas.071524598
- [65] McCook LJ, Jompa J, Diaz-Pulido G. Competition between corals and algae on coral reefs: A review of evidence and mechanisms. Coral Reefs. 2001;19:400-417
- [66] Kuffner IB, Paul V. Effects of the benthic cyanobacterium Lyngbya majuscula on larval recruitment of the reef corals Acropora surculosa and Pocillopora damicornis. Coral Reefs. 2004;23:455-458
- [67] Vermeij M, van Moorselaar I, Engelhard S, Hörnlein C, Vonk S, Visser P. The effects of nutrient enrichment and herbivore abundance on the ability of turf algae to overgrow coral in the Caribbean. PLoS One. 2010;5(12):e14312. DOI: 10.1371/journal. pone.0014312
- [68] Tan LT, Goh BPL. Chemical ecology of marine cyanobacterial secondary metabolites: A mini review. Journal of Coastal Development. 2009;13:1-9
- [69] Pennings SC, Weiss AM, Paul VJ. Secondary metabolites of the cyanobacterium Microcoleus lyngbyaceus and the sea hare Stylocheilus longicauda: Palatability and toxicity. Marine Biology. 1996;126:735-743
- [70] Thacker R, Nagle D, Paul VJ. Effects of repeated exposures to marine cyanobacterial secondary metabolites on feeding by juvenile rabbitfish and parrotfish. Marine Ecology Progress Series. 1997;147:21-29
- [71] Thacker RW, Becerro MA, Lumbang WA, Paul VJ. Allelopathic interactions between sponges on a tropical reef. Ecology. 1998;**79**:1740-1750
- [72] Nagle D, Paul VJ. Production of secondary metabolites by filamentous tropical marine cyanobacteria: Ecological functions of the compounds. Journal of Phycology. 1999;35:1412-1421
- [73] Nagle DG, Paul VJ, Roberts AM. Ypaoamide, a new broadly acting feeding deterrent from the marine cyanobacterium *Lyngbya majuscula*. Tetrahedron Letters. 1996;**37**:6263-6266

- [74] Nagle DG, Camacho FT, Paul VJ. Dietary preferences of the opisthobranch mollusc Stylocheilus longicauda for secondary metabolites produced by the tropical cyanobacterium Lyngbya majuscula. Marine Biology. 1998;132:267-273
- [75] Luesch H, Harrigan GG, Goetz G, Horgen FD. The cyanobacterial origin of potent anticancer agents originally isolated from sea hares. Current Medicinal Chemistry. 2002; 9:1791-1806
- [76] Díaz-Pulido G, McCook LJ. Macroalgae [Seaweeds]. In: Chin A, editor. The State of the Great Barrier Reef On-line. Townsville: Great Barrier Reef Marine Park Authority; 2008 http://www.gbrmpa.gov.au/corp_site/info_services/publications/sotr/downloads/ SORR_Macroalgae.pdf Accessed: 2017-03-30
- [77] Duffy JE, Hay ME. Seaweed adaptations to herbivory. Bioscience. 1990;40:368-376
- [78] Hay ME. Marine-terrestrial contrasts in the ecology of plant chemical defenses against herbivores. Trends in Ecology and Evolution. 1991;6:362-365
- [79] Hay ME, Kappel QE, Fenical W. Synergisms in plant defenses against herbivores: Interactions of chemistry, calcification, and plant quality. Ecology. 1994;75:1714-1726
- [80] Littler MM, Littler DS. Models of tropical reef biogenesis: The contribution of algae. Progress in Phycological Research. 1984;3:323-364
- [81] Heyward AJ, Negri AP. Natural inducers of coral larval metamorphosis. Coral Reefs. 1999;18:273-279
- [82] Littler MM, Littler DS, Brooks BL. Harmful algae on tropical coral reefs: Bottom-up eutrophication and top-down herbivory. Harmful Algae. 2006;5:565-585
- [83] Hughes TP. Catastrophes, phase shifts, and large-scale degradation of a Caribbean coral reef. Science. 1994;265:1547-1551
- [84] Mumby PJ, Steneck RS, Edwards AJ, Ferrari R, Coleman R, Harborne AR, Gibson PJ. Fishing down a Caribbean food web relaxes trophic cascades. Marine Ecology Progress Series. 2012;445:13-24
- [85] Littler MM, Littler DS, Brooks BL. Herbivory, nutrients, stochastic events, and relative dominances of benthic indicator groups on coral reefs: A review and recommendations. Smithsonian Contributions to the Marine Sciences. 2009;(38):401-414
- [86] McCook LJ. Macroalgae, nutrients and phase shifts on coral reefs: Scientific issues and management consequences for the Great Barrier Reef. Coral Reefs. 1999;18:357-367
- [87] Jompa J, McCook LJ. The effects of nutrients and herbivory on competition between a hard coral (Porites cylindrica) and a brown alga (Lobophora variegata). Limnology and Oceanography. 2002;47:527-534
- [88] Paerl H, Hall N, Calandrino E. Controlling harmful cyanobacterial blooms in a world experiencing anthropogenic and climatic-induced change. Science of the Total Environment. 2011;409:1739-1745. DOI: 10.1016/j.scitotenv.2011.02.001

- [89] Ahern KS, Ahern CR, Savige GM, Udy JW. Mapping the distribution, biomass and tissue nutrient levels of a marine benthic cyanobacteria bloom (*Lyngbya majuscule*). Marine and Fresh Water Research. 2007;**58**:883-904
- [90] Ahern KS, Ahern CR, Udy JW. Nutrient additions generate prolific growth of *Lyngbya majuscula* (cyanobacteria) in field and bioassay experiments. Harmful Algae. 2007; **6**:134-151
- [91] Thacker R, Paul VJ. Are benthic cyanobacteria indicators of nutrient enrichment? Relationships between cyanobacterial abundance and environmental factors on the reef flats of Guam. Bulletin of Marine Science. 2001;69:497-508
- [92] Thacker R, Paul VJ. Morphological, chemical, and genetic diversity of tropical marine cyanobacteria *Lyngbya* spp. and *Symploca* spp. (Oscillatoriales). Applied and Environmental Microbiology. 2004;**70**:3305-3312
- [93] Prato J. Afloramientos de cianobacterias marinas bentónicas en San Andrés, Providencia y las Islas del Rosario (Caribe colombiano): Caracterización y evaluación de su posible papel ecológico. [Master's Thesis]. Bogota: Universidad Nacional de Colombia; 2013
- [94] Heisler J, Glibert PM, Burkholder JM, Anderson DM, Cochlan W, Dennison WC, Dortch Q, Gobler CJ, Heil CA, Humphries E, Lewitus A, Magnien R, Marshall HG, Sellner K, Stockwell DA, Stoecker DK, Suddleson M. Eutrophication and harmful algal blooms: A scientific consensus. Harmful Algae. 2008;8:3-13
- [95] Watkinson AJ, JM O'N, Dennison WC. Ecophysiology of the marine cyanobacterium, *Lyngbya majuscula* (Oscillatoriaceae) in Moreton Bay, Australia. Harmful Algae. 2005;4:697-715
- [96] Osborne NJT, Webb PM, Shaw GR. The toxins of *Lyngbya majuscula* and their human and ecological health effects. Environment International. 2001;**27**:381-392
- [97] Paerl HW, Piehler MF. Nitrogen and marine eutrophication. In: Capone DG, Bronk DA, Mulholland MR, Carpenter EJ, editors. Nitrogen in the Marine Environment. 2nd ed. Oxford: Elsevier; 2008. p. 529-567. DOI: 10.1016/B978-0-12-372522-6.00011-6
- [98] Vivas-Aguas L, Tosic M, Sánchez J, Narváez S, Cadavid B, Bautista P, Betancourt J, Parra J, Echeverri L, Espinosa L. Diagnóstico y evaluación de la calidad ambiental marina en el Caribe y Pacífico colombiano. Red de vigilancia para la conservación y protección de las aguas marinas y costeras de Colombia-REDCAM. Informe técnico 2010. INVEMAR: Santa Marta; 2010. 208 p
- [99] O'Neil JM, Davis TW, Burford MA, Gobler CJ. The rise of harmful cyanobacteria blooms: The potential roles of eutrophication and climate change. Harmful Algae. 2012;14:313-334. DOI: 10.1016 j.hal.2011.10.027
- [100] Garzón-Ferreira J, Díaz JM. The Caribbean coral reefs of Colombia. In: Cortés J, editor. Latin American Coral Reefs. Amsterdam: Elsevier Science; 2003. p. 275-300

- [101] Aronson R, Precht W. Conservation, precaution, and Caribbean reefs. Coral Reefs. 2006;25:441-450. DOI: 10.1007/S00338-006-0122-9
- [102] Mora C. A clear human footprint in the coral reefs of the Caribbean. Proceedings of the Royal Society of London B. 2008;275:767-773. DOI: 10.1098/rspb.2007.1472
- [103] Logan A. Interespecific aggression in hermatypic corals from Bermuda. Coral Reefs. 1984;3:131-138
- [104] Lang JC, Chornesky EA. Competition between scleractinian reef corals: A review of mechanisms and effects. In: Dubinsky Z, editor. Coral Reefs. Amsterdam: Elsevier; 1990. p. 209-252
- [105] López-Victoria M, Zea S, Weil E. Competition for space between encrusting excavating Caribbean sponges and other coral reef organisms. Marine Ecology Progress Series. 2006;312:113-121
- [106] Chaves-Fonnegra A, Zea S. Coral colonization by the encrusting excavating Caribbean sponge Cliona delitrix. Marine Ecology. 2010;32:1-12. DOI: 10.1111/j.1439-0485.2010. 00416.xs
- [107] Chadwick NE, Morrow KM. Competition among sessile organisms on coral reefs. In: Dubinsky Z, Stambler N, editors. Coral Reefs: An Ecosystem in Transition. Heidelberg: Springer; 2011. p. 347-371
- [108] INVEMAR. 2005. Informe del los ambientes marinos y costeros en Colombia: Año 2005. Serie de publicaciones periódicas No. 8. INVEMAR: Santa Marta; 2005 360 p
- [109] Díaz-Pulido G, McCook LJ. The fate of bleached corals: Patterns and dynamics of algal recruitment. Marine Ecology Progress Series. 2002;232:115-128
- [110] Titlyanov E, Yakovleva I, Titlyanova T. Interaction between benthic algae (*Lyngbya bouil*lonii, Dictyota dichotoma) and scleractinian coral Porites lutea in direct contact. Journal of Experimental Marine Biology and Ecology. 2007;342:282-291
- [111] Golubic S, Abed RMM, Palinska K, Pauillac S, Chinain M, Laurent D. Marine toxic cyanobacteria: Diversity, environmental responses and hazards. Toxicon. 2010;56:836-841
- [112] Carpenter RC. Partitioning herbivory and its effects on coral reef algal comunities. Ecological Monographs. 1986;56:345-363
- [113] Hay ME, Fenical W. Marine plant-herbivore interactions: The ecology of chemical defense. Annual Review of Ecology and Systematics. 1988;19:111-145
- [114] Valentine JF, Heck KL, Kirsch KD, Webb D. Role of sea urchin Lytechinus variegatus grazing in regulating subtropical turtlegrass Thalassia testudinum meadows in the Florida Keys [USA]. Marine Ecology Progress Series. 2000;200:213-222
- [115] Hay ME. The role of seaweed chemical defenses in the evolution of feeding specialization and in the mediation of complex interactions. In: Paul VJ, editor. Ecological Roles for Marine Natural Products. Ithaca: Comstock Press; 1992. p. 93-118

- [116] Vaz Pinto F, Olabarria C, Arenas F. Role of top-down and bottom-up forces on the invasibility of intertidal macroalgal assemblages. Journal of Sea Research. 2013;**76**:178-186
- [117] Burkepile DE, Hay ME. 2006. Herbivore versus nutrient control of marine primary producers: Context-dependent effects. Ecology. 2006;87:3128-3139
- [118] Hay ME, Steinberg PD. The chemical ecology of plant-herbivore interactions in marine versus terrestrial communities. In: Rosenthal J, Berenbaum M, editors. Herbivores: Their Interaction with Secondary Metabolites, Evolutionary and Ecological Processes. San Diego: Academic Press; 1992. p. 371-413
- [119] Paul VJ, Hay ME. Seaweed susceptibility to herbivory: Chemical and morphological correlates. Marine Ecology Progress Series. 1986;33:255-264
- [120] Kiryu Y, Landsberg JH, Peters EC, Tichenor E, Burleson C, Perry N. Pathological effects of cyanobacteria on sea fans in southeast Florida. Journal of Invertebrate Pathology. 2015;129:13-27. DOI: 10.1016/j.jip.2015.04.007
- [121] Smith JE, Kuwabara J, Flanagan K, duPlessis S, Coney J, Beets J, Takabayashi M, Barnes S, Turner J, Brown D, Griesemer BK, Stanton F. An unusual cyanobacterial bloom in Hawai'i. Coral Reefs. 2008;27:851
- [122] Birrell CL, McCook LJ, Willis BL. Effects of algal turfs and sediment on coral settlement. Marine Pollution Bulletin. 2005;51:408-414
- [123] Diaz-Pulido G, Harii S, McCook LJ, Hoegh-Guldberg O. The impact of benthic algae on the settlement of a reef-building coral. Coral Reefs. 2010;29:203-208
- [124] Belov AP. A model of phycotoxin release by cyanobacterial cells. Ecological Modelling. 1998;**110**:105-117
- [125] Ross C, Santiago-Vazquez L, Paul V. Toxin release in response to oxidative stress and programmed cell death in the cyanobacterium *Microcystis aeruginosa*. Aquatic Toxicology. 2006;78:66-73
- [126] Preubel K, Wessel G, Fastner J, Chorus I. Response of cylindrospermopsin production and release in *Aphanizomenon flos-aquae* (Cyanobacteria) to varying light and temperature conditions. Harmful Algae. 2009;8:645-650
- [127] Nieto CF. Cianobacterias bentónicas arrecifales: Evaluación de sus interacciones alelopáticas mediante ensayos in situ en comunidades coralinas Islas del Rosario, Caribe Colombiano [Undergraduate thesis]. Bogota: Universidad Jorge Tadeo Lozano; 2012
- [128] Paerl HW, Gardner WS, Havens KE, Joyner AR, McCarthy MJ, Newell SE, Qin B, Scott JT. Mitigating cyanobacterial harmful algal blooms in aquatic ecosystems impacted by climate change and anthropogenic nutrients. Harmful Algae. 2016;54:213-222. DOI: /10.1016/j.hal.2015.09.009
- [129] Ainsworth CH, Mumby PJ. Coral-algal phase shifts alter fish communities and reduce fisheries production. Global Change Biology. 2015;21:165-172. DOI: 10.1111/gcb.12667

- [130] Diaz-Pulido G, Gouezo M, Tilbrook B, Dove SG, Anthony KRN. High CO, enhances the competitive strength of seaweeds over corals. Ecology Letters. 2010;14:156-162. DOI: 10.1111/j.1461-0248.2010.01565.x
- [131] Gardner TA, Cote IM, Gill JA, Grant A, Watkinson AR. Long-term region-wide declines in Caribbean corals. Science. 2003;301:958-960
- [132] Martinez S, Acosta A. Cambio temporal en la estructura de la comunidad coralina del área de Santa Marta-Parque Nacional Natural Tayrona [Caribe Colombiano]. Boletín de Investigaciones Marinas Y Costeras. 2005;34:161-191
- [133] Rodríguez-Ramírez A, Reyes-Nivia MC, Zea S, Navas-Camacho R, Garzón-Ferreira J, Bejarano S, Orozco C. Recent dynamics and condition of coral reefs in the Colombian Caribbean. Revista de Biología Tropical. 2010;58(Suppl 1):107-131
- [134] Schutte VGW, Selig ER, Bruno JF. Regional spatio-temporal trends in Caribbean coral reef benthic communities. Marine Ecology Progress Series. 2010;402:115-122
- [135] Zapata FA, Rodríguez-Ramírez A, Caro-Zambrano C, Garzón-Ferreira J. Mid-term coral-algal dynamics and conservation status of a Gorgona Island (Tropical Eastern Pacific) coral reef. Revista de Biología Tropical. 2010;58(Suppl 1):81-94
- [136] Lodeiros C, García N. The use of sea urchins to control fouling during suspended culture of bivalves. Aquaculture. 2004;231:293-298
- [137] Westbrook CE, Ringang RR, Cantero SMA, HDAR & TNC Urchin Team, Toonen RJ. Survivorship and feeding preferences among size classes of outplanted sea urchins, Tripneustes gratilla, and possible use as biocontrol for invasive alien algae. PeerJ. 2015; 3:e1235. DOI: 10.7717/peerj.1235
- [138] Burke L, Maidens J. Reefs at Risk in the Caribbean. Executive Summary. World Resources Institute: Washington; 2004. 16 p