

Prediction of the itraconazole minimal inhibitory concentrations (MICs) of chromoblastomycosis agents using Fourier Transform-Infrared Spectroscopy and chemometrics

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Objectives: use Fourier Transform-Infrared Spectroscopy (FTIR) and chemometrics to predict the itraconazole minimal inhibitory concentrations (MICs) for chromoblastomycosis (CBM) agents.

Methods: seventy-seven isolates of five genera of CBM agents were identified by comparison of ITS sequences of type strains available in GenBank using BLAST algorithm. Antifungal susceptibility of CBM agents to itraconazole (Sigma-Aldrich, USA) was performed according to the protocol M38-A2 of the Clinical and Laboratory Standards Institute (CLSI), utilizing the microdilution technique with a final antifungal concentration varying between 0.03-16 µg/mL to determine MICs. For FTIR analysis, the strains were prepared for Attenuated Total Reflection (ATR) with a new methodology using slices in glass fixing-modeling proposed. Five spectra were recorded from 4000 to 650 cm⁻¹ for each strain. The data set was sample pre-processed by amplitude normalization and 1st derivative (5 points) in Pirouette® Software. PLS-FTIR algorithm was performed with quintuplicates using two orthogonal signal correction components and leave-one-out cross-validation.

Results: the strains were identified with 99% of identity with type strain of each species of thirteen species distributed on five genera of CBM. The identified species were: *Fonsecaea pedrosoi* (41), *Fonsecaea monophora* (15), *Fonsecaea pugnacious* (1), *Fonsecaea nubica* (1), *Cladophialophora carrionii* (3), *Cladophialophora bantiana* (1), *Phialophora verrucosa* (2), *Phialophora americana* (5), *Exophiala spinifera* (3), *Exophiala xenobiotica* (2), *Rhinochadiella aquaspersa* (1), *Rhinochadiella tropicalis* (1) e *Rhinochadiella similis* (1). The geometric mean of itraconazole MICs was 0.784 µg/mL ranging between 0.06 µg/mL and 32.0 µg/mL, with MIC₅₀ equal to 0.5 µg/mL and MIC₉₀ equal to 2.0 µg/mL. FTIR analysis presented bands in the regions between 4000 and 3100 cm⁻¹ (attributed to NH and OH stretching) together with the bands 1650 (amide I of proteins), 1050 and 1025 (polysaccharides bands) and the 900-800 region (fingerprint) (**Figure 1**). In the PLS-FTIR model, the merit figures presented error less than 0.001 microgram/mL and determination coefficient (R²) of 1.00 with 10 latent variables (**Figure 2**).

The major contributions to regression model were observed in fingerprint region (regression vector, **Figure 1**).

Conclusion: CBM is subcutaneous mycosis caused by different dematiaceous fungus belonging to many species and genera. Considering that each species can show different responses to antifungals, it is fundamental to determine the MIC. This model enables direct prediction of MICs, without necessity to identify the CBM agent or to perform antifungals susceptibility assays, which are more expensive and laborious than the methodology proposed here.

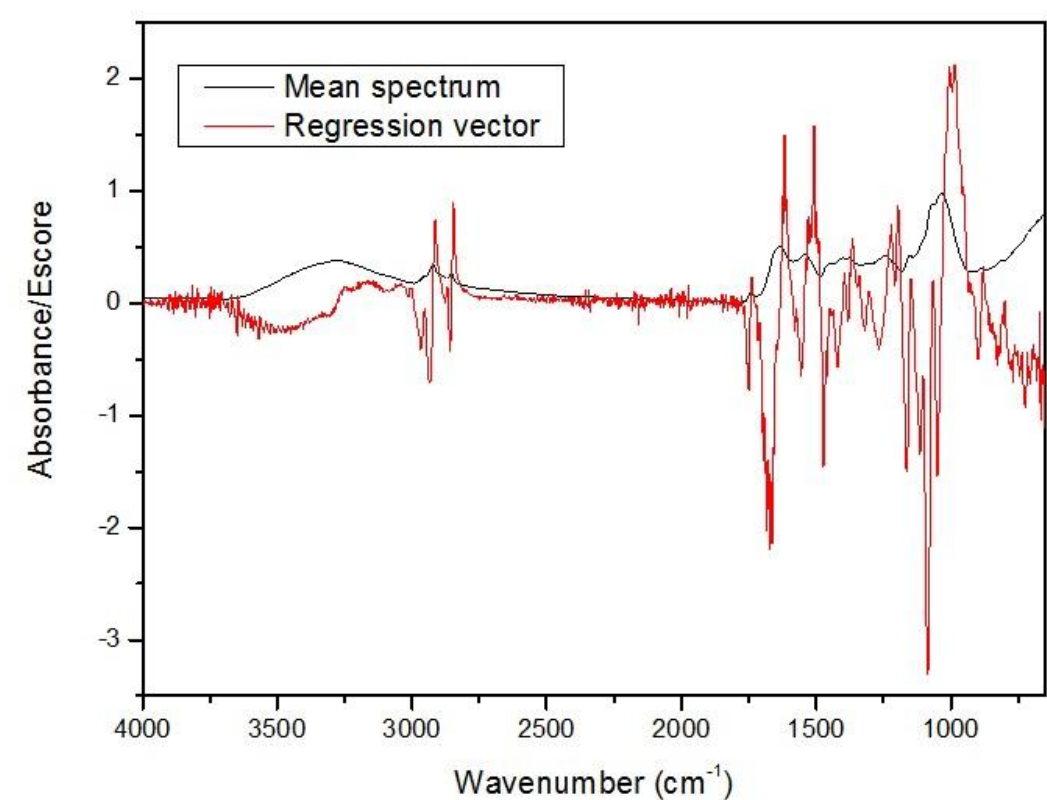


Figure 1. FT-IR mean spectrum of 77 samples (385 spectra) of chromoblastomycosis agents and regression vector of PLS-FTIR model of prediction of antifungal susceptibility to itraconazole supervised by CLSI method.

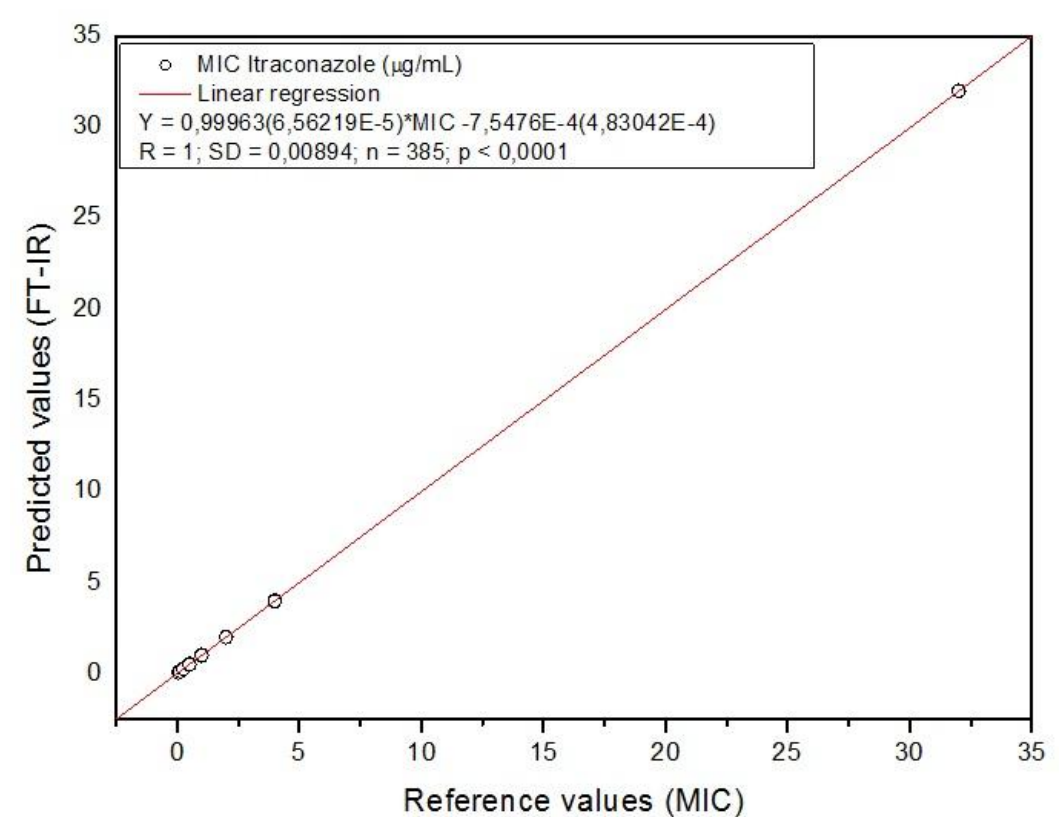


Figure 2. Regression analysis of PLS-FTIR model of prediction of MIC of 77 samples of chromoblastomycosis agents against itraconazole supervised by CLSI method.