Diagnostic accuracy of digital RNA quantification versus real-time PCR for the detection of respiratory syncytial virus in nasopharyngeal aspirates from children with acute respiratory infection

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Respiratory syncytial virus (RSV) is one of the most common etiological agents of acute respiratory infections (ARI) among children such as bronchiolitis and pneumonia.¹ Virusspecific molecular assays such as real-time polymerase chain reaction (RT-PCR) are now considered the gold standard in the diagnosis of viral respiratory tract infections, but simultaneous (multiplex) detection of different pathogens is limited which are considered majors limitations.² A multiplex digital method of RNA quantification, nCounter (NanoString Technologies), can overcome this disadvantage and identify, in a single reaction, the presence of different respiratory viruses.³ We aimed to evaluate the accuracy of nCounter (Nanostring Technologies) to identify and quantify RSV-A and RSV-B in NPA of children with ARI using real-time PCR as the reference method.

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NPA was collected at enrolment in a prospective cross-sectional study conducted in an Emergency Department from September 2009 to October 2013, in Salvador, Brazil. A quantitative RT-PCR with a subgroup-specific primer and probeset for RSVA and RSV-B was performed in parallel with a customized nCounter probeset containing viral targets in NPA.

Table 1A. Diagnostic accuracy of nCounter to detect RSV-A

		RT-PCR		Total
		Yes	No	
	Yes	55 (74.3%)	8 (1.6%)	63 (11.3%)
nCounter	No	19 (25.7%)	477 (98.4%)	496 (88.7%)
	Total	74 (13.2%)	485 (86.8%)	559 (100%)

Table 1B. Diagnostic accuracy of nCounter to detect RSV-B

		RT-PCR		Total
		Yes	No	
nCounter	Yes	52 (77.6%)	11 (2.2%)	63 (11.3%)
	No	15 (22.4%)	481 (97.8%)	496 (88.7%)
	Total	67 (12.0%)	492 (88.0%)	559 (100%)

This study group comprised 559 cases and RSV was detected by RT-PCR in 139 (24.9%) cases, RSV-A in 74 (13.2%) cases, and RSV-B in 67 (12.0%) cases. Two (1.4%) were coinfected.

Digital quantification by nCounter, the new diagnostic method, detected RSV in 122 (21.8%) samples, RSV-A in 63 (11.3%) and RSV-B in 63 (11.3%). Co-infections were detected in 4 (3.3%) cases. Interestingly, detection of RSV-A and RSV-B by any method occurred in 158 (28.3%) cases, both methods might be indicating complementary in detecting the complete **RSV** epidemic. The validation of nCounter as a qualitative (presence VS. absence), measure considering RT-PCR as the reference standard is shown in Table 1A for RSV-A and in Table 1B for RSV-B.

Overall, accuracy was 95.2% (95%CI:93.1%-96.7%) for RSV-A and 95.3% (95%CI:93.3%-96.9%) for RSV-B. Moreover, quantification of both RSV-A and RSV-B viral RNA was significantly correlated between nCounter and RT-PCR, as shown in Fig. 1A-B. Again, using RT-PCR as a reference, a significant correlation (Spearman r=0.44, p=8x10⁻⁵) was found in RSV-A-positive (n=74), between quantitative samples by nCounter (measured as detection normalized counts) and RT-PCR (measured as copies/ml). Similarly, in RSV-B RT-PCRpositive samples (n=67), a significant correlation (Spearman r=0.73, p= $3x10^{-12}$) was found between RSV-B quantitative detection by nCounter and RT-PCR (Fig. 1B



Figure 1: Correlation between RSV-A and RSV-**B** levels quantified by nCounter and Real-Time **PCR**. (A) In RT-PCR-positive samples (n=74), a significant correlation (Spearman r=0.44, p=8x10-5) was found between RSV-A quantitative detection by nCounter (measured as normalized counts) and RT-PCR (measured as copies/ml). (B) In RT-PCR-positive samples (n=67), a significant correlation (Spearman r=0.73, p=3x10-12) was found between RSV-B quantitative detection by nCounter (normalized counts) and RT-PCR (copies/ml).

In conclusion, digital RNA quantification of RSV-A and RSV-B by nCounter is highly accurate (>95%), using real-time PCR as a reference. Its robustness, high-throughput multiplex capacity and detection of cases undetected by real-time PCR indicate its suitability for large-scale epidemiological studies.